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# 表没食子儿茶素没食子酸酯抑制人腹膜间皮细胞 上皮-间质转化的作用及机制\*

谢斌<sup>1</sup>, 洪慧<sup>1</sup>, 帅欢<sup>1</sup>, 张林<sup>2△</sup>

(1. 湖南中医药高等专科学校附属第一医院, 湖南 株洲 412000; 2. 湖南省长沙市第四医院, 湖南 长沙 410006)

**摘要:**目的 探讨表没食子儿茶素没食子酸酯(EGCG)对人腹膜间皮细胞(HPMCs)上皮-间质转化(EMT)的作用及机制。方法 收集新开管的腹膜透析和腹膜透析1年以上患者的HPMCs,分别采用逆转录实时荧光定量聚合酶链反应(qRT-PCR)法和免疫印迹(Western blot)法检测紧密连接蛋白1(ZO-1)、E-钙黏蛋白(E-cadherin)、铁死亡抑制蛋白1(FSP1)mRNA和蛋白表达水平。采用HPMCs细胞系HMrSV5进行体外实验研究,分别加入20,40,60,80,100 μg/mL EGCG,检测EGCG对HMrSV5的细胞毒性。通过200 μg/mL和500 μg/mL晚期糖基化终产物(AGEs)构建HPMCs EMT模型,将HMrSV5细胞分为对照组(AGEs造模前无处理)、实验组(AGEs造模前2 h加入EGCG共孵育)、si-NC+EGCG组(转染si-NC后,AGEs造模前2 h加入EGCG共孵育)、si-Notch3+EGCG组(转染si-Notch3后,AGEs造模前2 h加入EGCG共孵育),检测ZO-1, E-cadherin, FSP1, Notch3 mRNA和蛋白表达水平。结果 与新开管的腹膜透析患者比较,腹膜透析1年以上患者HPMCs的ZO-1及E-cadherin mRNA和蛋白表达水平均显著升高, FSP1及Notch3 mRNA表达水平均显著降低( $P < 0.05$ )。与正常HMrSV5比较,加入500 μg/mL AGEs诱导HMrSV5后,ZO-1及E-cadherin mRNA和蛋白表达水平均显著升高, FSP1 mRNA和蛋白表达水平均显著降低( $P < 0.05$ )。与对照组比较,实验组ZO-1及E-cadherin mRNA和蛋白表达水平均显著降低, FSP1及Notch3 mRNA和蛋白表达水平均显著升高( $P < 0.05$ )。与si-NC+EGCG组比较,si-Notch3+EGCG组Notch3及FSP1 mRNA和蛋白表达水平均显著降低,ZO-1及E-cadherin mRNA和蛋白表达水平均显著升高( $P < 0.05$ )。腹膜透析患者HPMCs中Notch3表达水平与ZO-1和E-cadherin呈负相关( $r = -0.8477, -0.3822, P < 0.05$ ),与FSP1呈正相关( $r = 0.6626, P < 0.05$ )。结论 EGCG通过Notch3抑制AGEs诱导的HPMCs EMT,发挥对腹膜透析患者的保护作用。

**关键词:**表没食子儿茶素没食子酸酯;人腹膜间皮细胞;上皮-间质转化;晚期糖基化终产物;腹膜透析

## Effect and Mechanism of Epigallocatechin Gallate on Inhibiting Epithelial - Mesenchymal Transition of Human Peritoneal Mesothelial Cells

XIE Bin<sup>1</sup>, HONG Hui<sup>1</sup>, SHUAI Huan<sup>1</sup>, ZHANG Lin<sup>2</sup>

(1. The First Affiliated Hospital of Hunan Traditional Chinese Medical College, Zhuzhou, Hunan, China 412000; 2. The Fourth Hospital of Changsha, Changsha, Hunan, China 410006)

**Abstract: Objective** To investigate the effect and mechanism of epigallocatechin gallate (EGCG) on inhibiting the epithelial - mesenchymal transition (EMT) of human peritoneal mesothelial cells (HPMCs). **Methods** HPMC from patients starting peritoneal

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第一作者:谢斌,女,硕士研究生,主治医师,研究方向为慢性肾脏病的诊疗,(电子信箱)sai158991@163.com。

△通信作者:张林,男,博士研究生,副主任医师,研究方向为血液净化,(电子信箱)397639438@qq.com。

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dialysis (PD) and underwent PD for more than one year were collected, and the expression levels of zonula occluden - 1 (ZO - 1), E - cadherin and ferroptosis suppressor protein 1 (FSP1) mRNA and protein were detected by the reverse transcription real - time fluorescence quantitative polymerase chain reaction (qRT - PCR) and Western blot, respectively. HPMCs cell line HMrSV5 was used for *in vitro* study, and 20, 40, 60, 80, 100  $\mu\text{g}/\text{mL}$  EGCG were added into HMrSV5 cell culture fluid respectively to detect the cytotoxicity of EGCG to HMrSV5 cells. The EMT model of HPMCs was constructed by 200  $\mu\text{g}/\text{mL}$  and 500  $\mu\text{g}/\text{mL}$  advanced glycation end products (AGEs). HMrSV5 cells were divided into the control group (no treatment before modeling by AGEs), the test group (EGCG was added to co - incubate with HMrSV5 cells 2 h before modeling by AGEs), the si - NC + EGCG group (EGCG was added to co - incubate with HMrSV5 cells after transfection of si - NC and 2 h before modeling by AGEs) and the si - Notch3 + EGCG group (EGCG was added to co - incubate with HMrSV5 cells after transfection of si - Notch3 and 2 h before modeling by AGEs). The expression levels of ZO - 1, E - cadherin, FSP1, Notch3 mRNA and protein were detected.

**Results** Compared with those in the patients starting PD, the expression levels of ZO - 1 and E - cadherin mRNA and protein of HPMCs in patients underwent PD for more than one year were significantly higher, and the expression levels of FSP1 and Notch3 mRNA of HPMCs in patients underwent PD for more than one year were significantly lower ( $P < 0.05$ ). Compared with those in the normal HMrSV5 cells, the expression levels of ZO - 1 and E - cadherin mRNA and protein in the HMrSV5 cells induced by 500  $\mu\text{g}/\text{mL}$  AGEs were significantly higher, and the expression levels of FSP1 mRNA and protein in the HMrSV5 cells induced by 500  $\mu\text{g}/\text{mL}$  AGEs were significantly lower ( $P < 0.05$ ). Compared with those in the control group, the expression levels of ZO - 1 and E - cadherin mRNA and protein in the test group were significantly lower, the expression levels of FSP1 and Notch3 mRNA and protein in the test group were significantly higher ( $P < 0.05$ ). Compared with those in the si - NC + EGCG group, the expression levels of Notch3 and FSP1 mRNA and protein in the si - Notch3 + EGCG group were significantly lower, and the expression levels of ZO - 1 and E - cadherin mRNA and protein in the si - Notch3 + EGCG group were significantly higher ( $P < 0.05$ ). The Notch3 expression level of HPMCs in patients underwent PD was negatively correlated with the expression levels of ZO - 1 and E - cadherin ( $r = -0.8477$  and  $-0.3822$ ,  $P < 0.05$ ), while positively correlated with the expression level of FSP1 ( $r = 0.6626$ ,  $P < 0.05$ ). **Conclusion** EGCG can play a protective role for patients underwent PD by inhibiting AGEs - induced EMT of HPMCs through Notch3.

**Key words:** epigallocatechin gallate; human peritoneal mesothelial cells; epithelial - mesenchymal transition; advanced glycation end products; peritoneal dialysis

腹膜透析(PD)是终末期肾病的替代疗法,占全球所有形式肾病替代治疗的10%以上<sup>[1]</sup>。上皮 - 间质转化(EMT)是上皮细胞在高糖、转化生长因子 -  $\beta$ (TGF -  $\beta$ )等因素诱导下,上皮细胞特异性标志物表达丧失,并获得肌成纤维细胞表型的过程<sup>[2]</sup>。人腹膜间皮细胞(HPMCs)被认为是腹膜纤维化的核心机制<sup>[3]</sup>。腹膜纤维化的发生会导致PD失败,极大地限制了PD的应用。绿茶提取物表没食子儿茶素没食子酸酯(EGCG)是绿茶多酚的主要单体及活性成分,其化学结构中包含没食子酰侧链,具有抗氧化、抗肿瘤、抗炎等生物学特性<sup>[4]</sup>。最新的研究发现,EGCG可通过增加细胞内自噬而抑制人成纤维细胞的肌成纤维细胞转化<sup>[5]</sup>。PANJI等<sup>[6]</sup>研究发现,EGCG通过活性氧(ROS)/Smad信号传导抑制TGF -  $\beta$ 诱导的人宫颈癌细胞EMT。Notch是编码哺乳动物细胞中跨膜受体家族的一组基因,主要调控细胞的增殖、分化和凋亡,影响细胞的正常生长,在机体稳态、血管生成和癌症发生中起重要作用,Notch受体家族有4个成员,其中Notch受体3(Notch3)较常见<sup>[7]</sup>。ZHU等<sup>[8]</sup>的研究结果显示,EGCG通过靶向抑制Notch信号通路,抑制链脲佐菌素诱导的糖尿病模型小鼠的肾纤维化过程,提示EGCG可能是一种潜在的抗纤维化试剂,

可调节细胞EMT过程。本研究中通过体外建立晚期糖基化终产物(AGEs)诱导的HPMCs EMT的模型,并采用绿茶提取物EGCG对EMT过程进行干预,探讨EGCG对保护腹膜超滤功能的作用机制。现报道如下。

## 1 材料与方法

### 1.1 样本来源

腹膜来源于新开管的PD和PD 1年以上患者,获取的腹膜标本立即放入D - Hank液中低温保存,短时间内处理。所有患者均签署知情同意书,本研究方案经湖南中医药大学高等专科学校附属第一医院医学伦理委员会审批。

### 1.2 细胞与试剂

细胞:HPMCs细胞株HMrSV5(中国科学院上海细胞库,批号为XY - XB - 1663)。

试剂:EGCG(上海源叶公司,批号为B20106);AGEs(美国Biovision公司,批号为4271 - 100);Dulbecco改良Eagle培养基(DMEM,批号为SH30243),牛血清白蛋白(FBS,批号为SH30070.03),均购自美国Hyclone公司;Ham's F12培养基(批号为31765035),Lipofectamine™ 2000(批号为11668019),均购自美国Thermo Fisher公司;逆转录实时荧光定量聚合酶链反应(qRT - PCR)试剂盒(RNA提取试剂,批号为R401 - 01),逆转

录试剂盒(批号为 R223-01), SYBR Green 试剂(批号为 Q221-01), CCK-8 试剂盒(批号为 A311-01), 均购自南京诺唯赞生物科技股份有限公司; 紧密连接蛋白 1 (ZO-1) 抗体(批号为 ab276131), E-钙黏蛋白(E-cadherin) 抗体(批号为 ab40772), 铁死亡抑制蛋白 1 (FSP1) 抗体(批号为 ab197896), Notch3 抗体(批号为 ab300527), 均购自英国 Abcam 公司; si-NC, si-Notch3 (美国 Invitrogen 公司); ZO-1, E-cadherin, FSP1, Notch3 引物, 甘油醛-3-磷酸脱氢酶(GAPDH) 引物, 均购自生工生物工程(上海)公司。

### 1.3 方法

原代细胞提取和培养: 将无菌腹膜组织移入烧杯中, 加入预冷的磷酸盐缓冲液(PBS)中洗涤 2 次, 剥离多余的脂肪组织及血管, PBS 冲洗至无油珠漂浮, 将网膜组织平铺于无菌培养皿, 剪成 0.7 cm × 0.7 cm 大小, 加入胰蛋白酶和 0.02% 依地酸二钠(EDTA)消化液, 用吸管反复吹打均匀, 移入细胞培养瓶内消化 25 min, 每 5 min 振荡 1 次, 消化结束后, 将消化液经 100 目滤网过滤于烧杯中, 加入 Ham's F12 完全培养基终止消化, 离心, 收集细胞, 将细胞铺至细胞培养皿中, 在 5% CO<sub>2</sub>、37 °C 恒温培养箱中培养, 得到原代 HPMCs。HMrSV5 用含 15% FBS 的 DMEM, 在 5% CO<sub>2</sub>、37 °C 恒温培养箱中培养, 细胞密度达 80% 时用胰酶消化细胞, 显微镜下观察到大部分细胞变圆时终止消化, 加入新鲜培养基, 每隔 2 d 更换 1 次新鲜培养基, 待细胞生长密度达 80% 时进行消化和传代, 取对数生长期的细胞进行实验。

AGEs 诱导的 HPMCs EMT: AGEs 诱导 HMrSV5 的 EMT 模型, 将细胞分为 1 组(正常 HMrSV5, 培养基不加 AGEs)、2 组(加入 200 μg/mL AGEs 诱导 HMrSV5)、3 组(加入 500 μg/mL AGEs 诱导 HMrSV5)<sup>[9]</sup>, 培养 72 h 后收集细胞, 检测 ZO-1, E-cadherin, FSP1 的 mRNA 和蛋白表达水平。

qRT-PCR 法检测 HPMCs 中的 ZO-1, E-cadherin, FSP1, Notch3 mRNA 表达水平及相关性: TRIzol 试剂提取原代 HPMCs 和 HMrSV5 中的总 RNA, 酶标仪检测总 RNA 的纯度和含量, 按 qRT-PCR 试剂盒说明, 依次进行逆转录和 qRT-PCR 实验。qRT-PCR 反应体系为 cDNA 模板 2 ng, 上下游引物各 0.4 μL, SYBR Primix Ex TaqTM 5 μL, ddH<sub>2</sub>O 补充至 10 μL; qRT-PCR 反应条件为预变性 95 °C(30 s), 变性 95 °C(7 s), 退火 55 °C(30 s), 延伸 72 °C(15 s), 40 个循环周期。扩增结果根据 2<sup>-ΔΔCt</sup> 法计算 mRNA 的相对表达量, 采用 Spearman 相关性分析 ZO-1, E-cadherin, FSP1, Notch3 mRNA 表达的相关性。qRT-PCR 引物序列见表 1。

表 1 逆转录实时荧光定量聚合酶链反应引物序列

Tab. 1 Primer sequences of qRT-PCR

引物	正向	反向
GAPDH	5'-AATGGCAGCCGTTAGGAAA-3'	5'-GCGCCCAATACGACCAAATC-3'
ZO-1	5'-TGCTCTGTTGCCACTGTT-3'	5'-TCTGTACATGCTGGCCAAAG-3'
E-cadherin	5'-AAGTTGAGCCCAAGGTGAT-3'	5'-CTGGAAGGAGCGGTCTTTTT-3'
FSP1	5'-TCTTGGTTTATCCTGACTGCT-3'	5'-GCTGCTTATCTGGGAAGCCT-3'
Notch3	5'-CCAGCATCACCTGCCTGTTA-3'	5'-CCAAGTCTGACGTCCCTCAC-3'

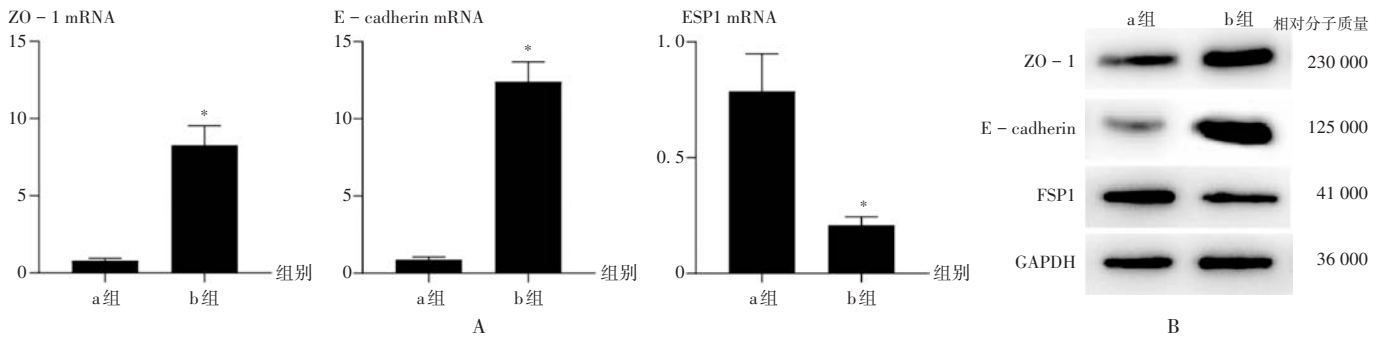
免疫印迹(Western blot)法检测 HPMCs 中 ZO-1, E-cadherin, FSP1, Notch3 的蛋白表达水平: 收集各组细胞, 使用 RIPA 裂解提取总蛋白, 通过 BCA 法进行蛋白定量。样品变性后, 通过十二烷基硫酸钠-聚丙烯酰胺凝胶电泳(SDS-PAGE)分离等量蛋白, 转移至聚偏二氟乙烯(PVDF)膜上, 5% 脱脂牛奶 4 °C 下封闭 1 h, TBST 洗膜 5 min, 共 3 次, 加入 ZO-1 抗体(1:1 000)、E-cadherin 抗体(1:1 000)、FSP1 抗体(1:1 000)、Notch3 抗体(1:1 000), 4 °C 摇床孵育过夜, TBST 洗膜 5 min, 共 3 次, 加入辣根过氧化物酶(HRP)标记的羊抗兔的二抗(1:1 000), 37 °C 下孵育 1 h, TBST 洗膜 5 min, 共 5 次。显影曝光, 利用 Image-Pro Plus 图像分析系统分析蛋白条带的灰度值。重复 3 次, 取平均值。

CCK-8 法检测 EGCG 对 HPMCs 的细胞毒性: 收集 HMrSV5, 用含 10% 胎牛血清的 DMEM 完全培养基稀释成单细胞悬液, 以 3 × 10<sup>4</sup> / mL 密度接种于 96 孔板, 接种体积为 200 μL。EGCG 溶于 DMEM 完全培养基中, 分别采用 0, 20, 40, 60, 80, 100 μg/mL EGCG 处理各组细胞, 在细胞培养箱中培养 24 h 后加入 20 μL CCK-8 检测试剂, 1 h 后用酶标仪于 450 nm 波长处测定吸光度。

细胞转染: 将 HMrSV5 细胞分为 si-NC + EGCG 组和 si-Notch3 + EGCG 组, 按美国 Invitrogen 公司 Lipofectamine 2000 说明书步骤进行转染。调整 HPMCs 数量, 接种于 6 孔板, 使用不含抗菌药物的完全培养基培养, 使转染时细胞生长密度达 70%。制备 Notch3 特异性 si-RNA (si-Notch3) 与脂质体复合物, 同时制备无干扰作用的 si-RNA (si-Control) 与脂质体复合物, 将复合物加入 6 孔板相应 si-Notch3 + EGCG 组和 si-NC + EGCG 组的孔内, 摇动、混匀, 6 h 后弃去细胞培养基, 用 PBS 洗涤细胞 2 次, 更换为 DMEM 完全培养基, 24 h 后更换为含有抗菌药物的完全培养基。采用 qRT-PCR 法检测转染后细胞中 Notch3 的表达。

### 1.4 统计学处理

采用 SPSS 20.0 和 GraphPad Prism 7.0 统计学软件分析。符合正态分布的计量资料以  $\bar{x} \pm s$  表示, 组间两两比较采用 LSD-t 检验, 多组间比较采用单因素方差分



注:a组为新开管PD患者组,b组为PD 1年以上患者组。与a组比较,\* $P < 0.05$ 。图5同。

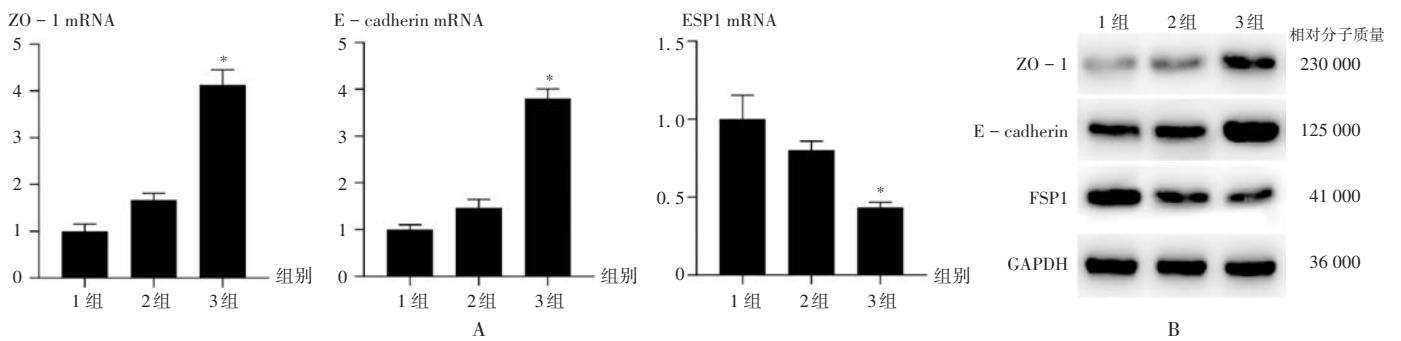
A. qRT-PCR法检测ZO-1,E-cadherin,FSP1 mRNA表达水平 B. Western blot法检测ZO-1,E-cadherin,FSP1 蛋白表达水平

图1 新开管PD和PD 1年以上患者HPMCs中ZO-1,E-cadherin,FSP1的mRNA和蛋白表达水平比较

Note:Group a refers to the group starting PD,and group b refers to the group underwent PD for more than one year. Compared with those in the group a,\* $P < 0.05$  (for Fig. 1 and Fig. 5).

A. Expression levels of ZO-1,E-cadherin and FSP1 mRNA (qRT-PCR) B. Expression levels of ZO-1,E-cadherin and FSP1 protein (Western blot)

Fig. 1 Comparison of expression levels of ZO-1,E-cadherin and FSP1 mRNA and protein of HPMCs in patients starting PD and underwent PD for more than one year



注:1组为正常HMrSV5,2组为200  $\mu\text{g}/\text{mL}$  AGEs诱导HMrSV5,3组为500  $\mu\text{g}/\text{mL}$  AGEs诱导HMrSV5。与1组比较,\* $P < 0.05$ 。

A. qRT-PCR法检测ZO-1,E-cadherin,FSP1 mRNA表达水平 B. Western blot法检测ZO-1,E-cadherin,FSP1 蛋白表达水平

图2 不同质量浓度AGEs诱导HPMCs EMT

Note:Group 1 refers to normal HMrSV5,group 2 refers to HMrSV5 induced by 200  $\mu\text{g}/\text{mL}$  AGEs,and group 3 refers to HMrSV5 induced by 500  $\mu\text{g}/\text{mL}$  AGEs. Compared with those in the group 1,\* $P < 0.05$ .

A. Expression levels of ZO-1,E-cadherin and FSP1 mRNA (qRT-PCR) B. Expression levels of ZO-1,E-cadherin and FSP1 protein (Western blot)

Fig. 2 EMT of HPMCs induced by different mass concentrations of AGEs

析。 $P < 0.05$ 为差异有统计学意义。

## 2 结果

### 2.1 不同阶段PD患者HPMCs中ZO-1,E-cadherin,FSP1表达水平比较

qRT-PCR法检测结果显示,与a组比较,b组患者的ZO-1和E-cadherin mRNA表达水平均显著升高,FSP1 mRNA表达水平显著降低( $P < 0.05$ ),详见图1 A。Western blot法检测结果显示,与a组比较,b组患者的ZO-1和E-cadherin蛋白表达水平均升高,FSP1蛋白表达水平降低,详见图1 B。结果表明,PD 1年患者的HPMCs已向EMT转化。

### 2.2 AGEs诱导HPMCs EMT

qRT-PCR法检测结果显示,与1组比较,3组患

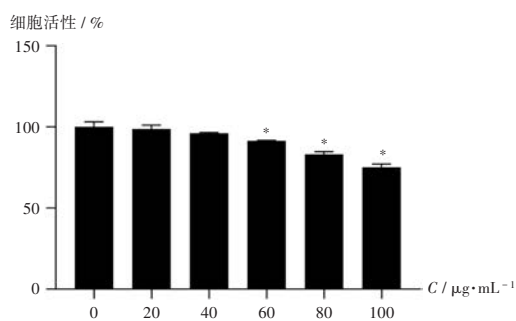
者的ZO-1和E-cadherin mRNA表达水平均显著升高,FSP1 mRNA表达水平显著降低( $P < 0.05$ ),详见图2 A。Western blot法检测结果显示,加入500  $\mu\text{g}/\text{mL}$  AGEs诱导HMrSV5后,ZO-1和E-cadherin蛋白表达水平均升高,FSP1蛋白表达水平降低,详见图2 B。结果表明,500  $\mu\text{g}/\text{mL}$  AGEs可有效诱导HPMCs EMT,后续实验可选择此浓度构建EMT模型。

### 2.3 EGCG对HPMCs的细胞毒性

CCK-8法检测结果显示,与0  $\mu\text{g}/\text{mL}$ 比较,60,80,100  $\mu\text{g}/\text{mL}$  EGCG能显著抑制细胞的活性( $P < 0.05$ ),详见图3。故选择40  $\mu\text{g}/\text{mL}$ 为EGCG给药浓度。

### 2.4 Notch3在EGCG治疗HPMCs EMT中的作用

qRT-PCR法检测结果显示,与对照组比较,实验组



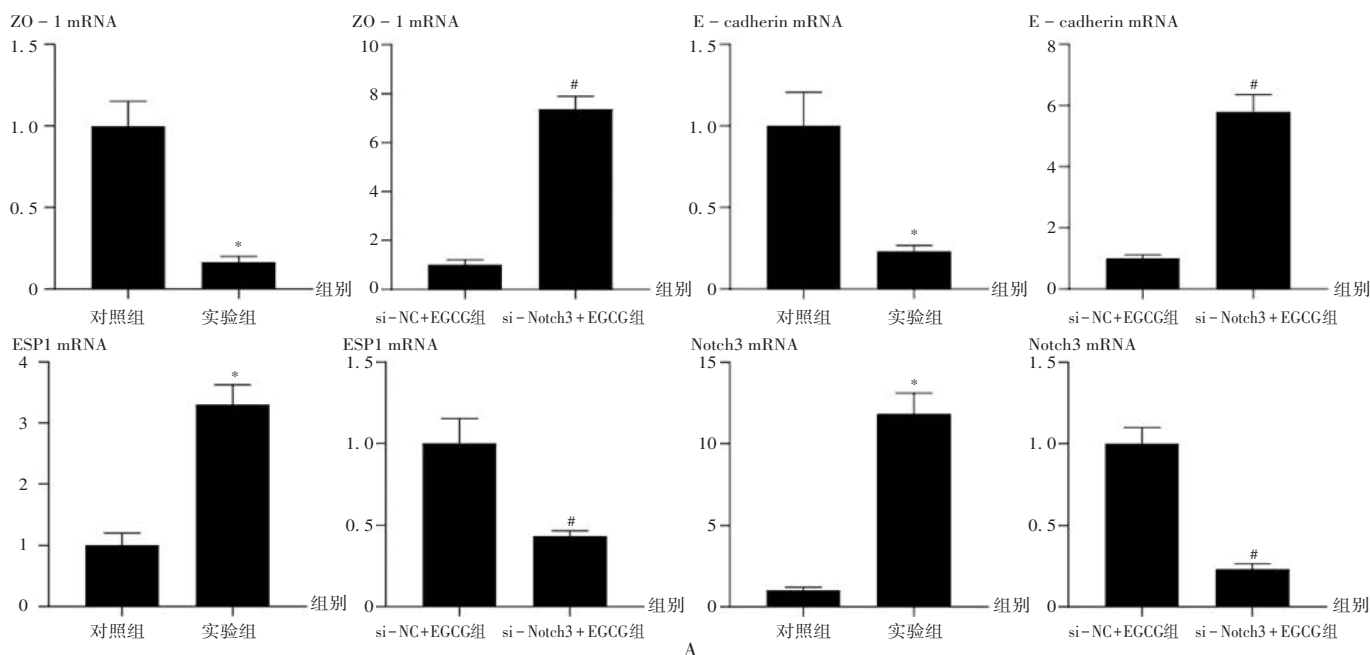
注:与0 μg/mL比较,\* $P < 0.05$ 。

图3 不同质量浓度EGCG对HMrSV5的细胞毒性

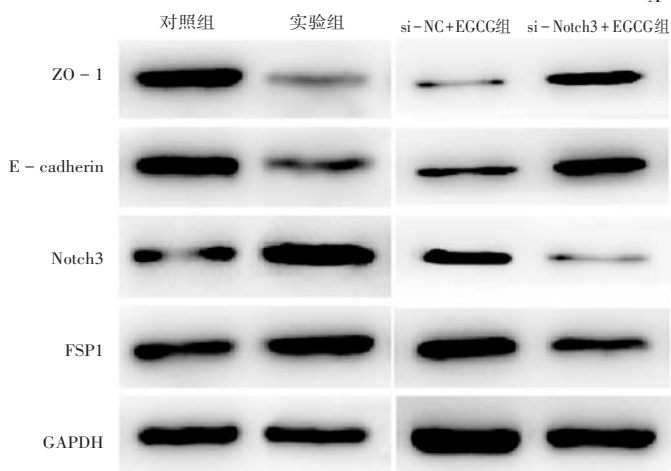
Note: Compared with those in the 0 μg/mL, \* $P < 0.05$ .

Fig. 3 Cytotoxicity of EGCG with different mass concentrations

on HMrSV5



A



B

注:与对照组比较,\* $P < 0.05$ ;与si-NC+EGCG组比较,# $P < 0.05$ 。

A. qRT-PCR法检测 B. Western blot法检测

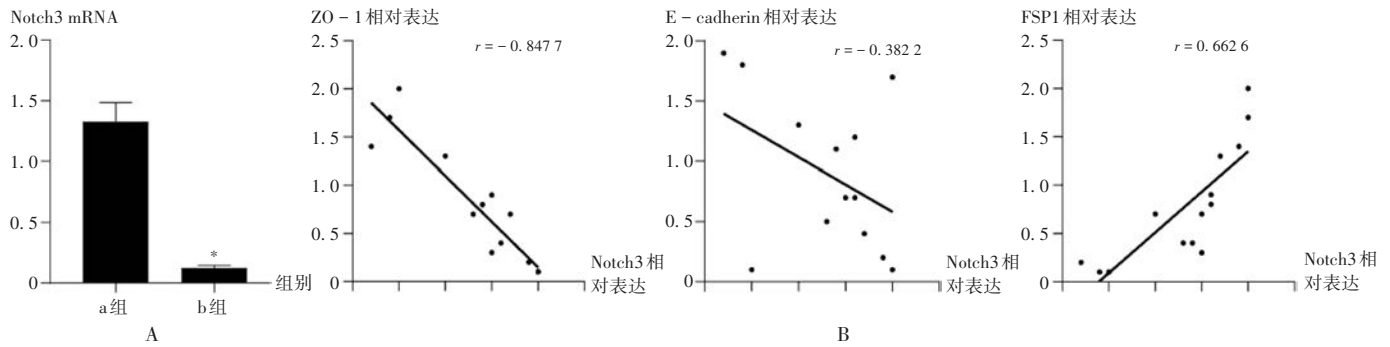
图4 Notch3对经EGCG干预的HPMCs中ZO-1,E-cadherin,FSP1 mRNA和蛋白表达水平的影响

Note: Compared with those in the control group, \* $P < 0.05$ ; Compared with those in the si-NC+EGCG group, # $P < 0.05$ .

A. Results of detection (qRT-PCR) B. Results of detection (Western blot)

Fig. 4 Effect of Notch3 on the expression levels of ZO-1,E-cadherin,FSP1 mRNA and protein in HPMCs after the intervention of EGCG

患者的ZO-1和E-cadherin mRNA表达水平均显著降低,FSP1和Notch3 mRNA表达水平均显著升高( $P < 0.05$ );与si-NC+EGCG组比较,si-Notch3+EGCG组患者的ZO-1和E-cadherin mRNA表达水平均显著升高,FSP1和Notch3 mRNA表达水平均显著降低( $P < 0.05$ ),详见图4 A。Western blot法检测结果显示,与对照组比较,实验组患者的ZO-1和E-cadherin蛋白表达水平均降低,FSP1和Notch3蛋白表达水平均升高;与si-NC+EGCG组比较,si-Notch3+EGCG组患者的ZO-1和E-cadherin蛋白表达水平均升高,FSP1和Notch3蛋白表达水平均降低,详见图4 B。



A. qRT-PCR法检测 Notch3 mRNA 表达水平 B. PD 患者中 Notch3 与 ZO-1, E-cadherin, FSP1 相关性分析

图5 PD患者HPMCs中Notch3与ZO-1, E-cadherin, FSP1表达水平相关性分析

A. Expression level of Notch3 mRNA (qRT-PCR) B. Correlation analysis of Notch3 with ZO-1, E-cadherin and FSP1 in patients underwent PD

Fig. 5 Correlation analysis of Notch3 with ZO-1, E-cadherin and FSP1 expression levels of HPMCs in patients underwent PD

## 2.5 PD患者HPMCs中Notch3与EMT基因表达水平相关性

采用qRT-PCR法检测PD患者HPMCs中Notch3 mRNA表达水平,结果显示,与a组比较,b组患者Notch3 mRNA表达水平显著降低( $P < 0.05$ ),详见图5A。Notch3表达水平与上皮分子标志物(ZO-1和E-cadherin)及间质分子标志物(FSP1)相关性分析结果显示,PD患者HPMCs中Notch3与ZO-1, E-cadherin的表达水平呈负相关( $r = -0.8477, -0.3822, P < 0.05$ ),与FSP1的表达水平呈正相关( $r = 0.6626, P < 0.05$ ),详见图5B。

## 3 讨论

我国慢性肾脏病发病率为10.8%<sup>[10]</sup>,部分患者进展至终末期肾病,PD已成为终末期肾病患者肾脏替代疗法的重要选择之一,但PD液的低pH、高葡萄糖、乳酸和葡萄糖降解产物会引起HPMCs的慢性炎症和损伤,导致腹膜纤维化而致PD失败<sup>[11]</sup>。EMT是HPMCs腹膜纤维化的核心机制。EMT是指上皮细胞通过特定程序转化为具有间质表型细胞的生物学过程<sup>[12]</sup>,在胚胎发育、慢性炎症、组织重建、癌症转移等多种纤维化疾病中起重要作用,主要特征为细胞黏附分子如E-cadherin表达下调、细胞角蛋白转化为波形蛋白细胞骨架,且形态学上具有外间充质细胞特点等<sup>[13]</sup>。

研究显示,高渗透压、高糖及各种细胞因子等刺激均可促进HPMCs EMT,使腹膜间皮细胞失去细胞极性和丧失黏附性,E-cadherin表达水平降低,向成纤维细胞转化,迁移和侵袭能力增强, $\alpha$ -平滑肌肌动蛋白( $\alpha$ -SMA)和波形蛋白过度表达等,HPMCs EMT是腹膜纤维化的起始及关键环节,且与腹膜功能密切相关<sup>[14]</sup>。因此,抑制HPMCs EMT对治疗PD具有重要意义。本研究中发现,相比于新开管的PD患者,PD1年以上患者HPMCs的ZO-1和E-cadherin表达水平均显著升高,FSP1表达水平显著降低,表明HPMCs已向EMT转化。AGEs能引

起体外培养的HPMCs形态逐渐变为梭形,上调HPMCs的 $\alpha$ -SMA表达,下调E-cadherin表达,从而诱导HPMCs上皮-间叶转化<sup>[15]</sup>。考虑到原代HPMCs难以转染等因素,本研究中采用HPMCs细胞系HMrSV5进行体外实验研究。结果显示,500  $\mu\text{g}/\text{mL}$  AGEs可显著上调HMrSV5细胞ZO-1和E-cadherin表达,显著下调FSP1表达。故选择500  $\mu\text{g}/\text{mL}$  AGEs作为造模剂量。绿茶提取物EGCG具有很强的生物活性和抗肾癌作用,能抑制肾癌细胞生长,缩小新生微血管面积<sup>[16]</sup>。在非酒精性脂肪性肝病模型小鼠中,EGCG治疗使肝纤维化相关基因显著下调,促进脂质和糖代谢、抗脂质过氧化和抗炎活性、抗纤维化,从而减缓非酒精性脂肪性肝病的进展<sup>[17]</sup>。本研究中,EGCG可显著减弱AGEs对ZO-1, E-cadherin, FSP1表达的影响,表明EGCG可抑制AGEs诱导的HPMCs EMT;通过抑制E-cadherin和波形蛋白的表达来抑制甲状腺癌细胞的EMT、侵袭和迁移,这可能通过调节TGF- $\beta$ /Smad信号通路实现<sup>[18]</sup>。此外,EGCG预防肾小管细胞EMT的分子机制很可能是通过激活转录因子核因子E2相关因子2(Nrf2)信号传导和增加过氧化氢酶抗氧化酶来减少细胞内ROS的产生<sup>[19]</sup>,但EGCG影响HPMCs EMT的具体作用机制仍不明确。

Notch3是Notch信号通路中的关键受体,可影响细胞的增殖、分化、迁移、生长、凋亡等过程。研究发现,赖氨酸特异性去甲基化酶1通过Jagged-1/Notch信号通路诱导EMT并促进肾纤维化<sup>[20]</sup>。MATSUURA等<sup>[21]</sup>研究发现,Notch3可能通过调节EMT来控制食管鳞状细胞癌的化疗敏感性。故本研究中重点关注Notch3在EGCG治疗HPMCs EMT中的作用。结果显示,EGCG可上调HPMCs中Notch3的表达,抑制Notch3表达可减弱EGCG对HPMCs中ZO-1和E-cadherin表达上调及FSP1表达下调的作用,表明EGCG对HPMCs EMT影响与调节Notch3表达有关。LIN等<sup>[22]</sup>研究发现,Notch3的

表达与GATA结合蛋白3(GATA-3)的表达呈正相关,Notch3胞内结构域的过表达可能通过激活GATA-3转录来抑制EMT的发展。另有研究表明,Notch3胞内结构域可激活雌激素受体 $\alpha$ (ER $\alpha$ )的表达,抑制乳腺癌细胞的EMT表型<sup>[21]</sup>。本研究中发现,PD患者HPMCs中Notch3与上皮分子标志(ZO-1和E-cadherin)表达呈负相关,与间皮分子标志物(FSP1)表达呈正相关,表明Notch3在PD患者HPMCs EMT中发挥重要作用。

综上所述,EGCG可抑制HPMCs EMT,其作用机制与调节Notch3表达有关。初步明确EGCG对HPMCs EMT的治疗作用,可为进一步研究EGCG对肾脏疾病的治疗提供参考。

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